

MECHANISTIC ASPECTS OF COENZYME B₁₂-DEPENDENT REARRANGEMENTS.
 ORGANOMETALLICS AS FREE RADICAL PRECURSORS

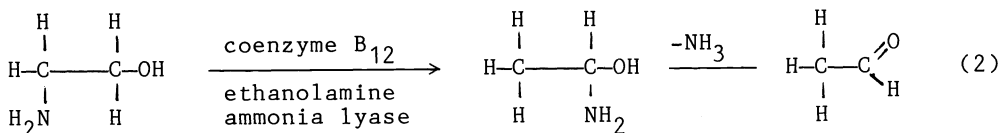
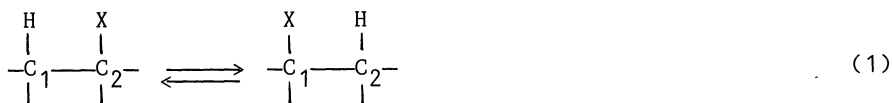
Jack Halpern

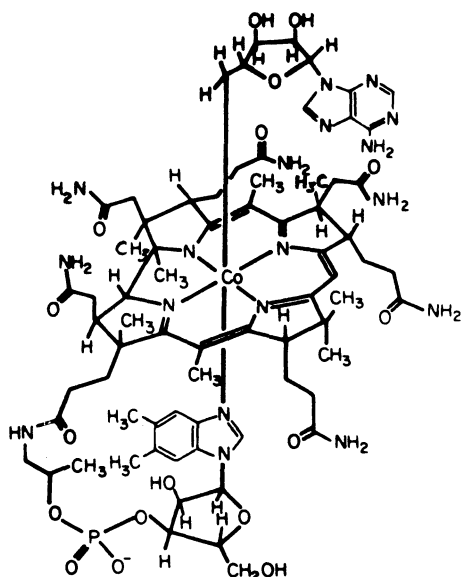
Department of Chemistry, The University of Chicago
 Chicago, Illinois 60637, U.S.A.

Abstract - The mechanistic aspects of coenzyme B₁₂-dependent rearrangements are discussed. At the present stage it appears that the principal, if not the only, role of coenzyme B₁₂ is to serve as the precursor for an organic free radical, generated by homolytic dissociation of the cobalt-carbon bond, which triggers the substrate rearrangement. The determination of cobalt-alkyl bond dissociation energies in organo-cobalt compounds related to coenzyme B₁₂, and the electronic and structural factors that influence such bond dissociation energies, are discussed. An analogy is developed between the role of coenzyme B₁₂ in biological systems as a reversible "free radical carrier" and the role of hemes as reversible dioxygen carriers.

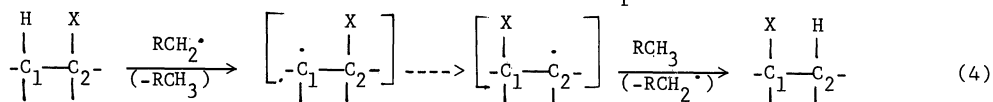
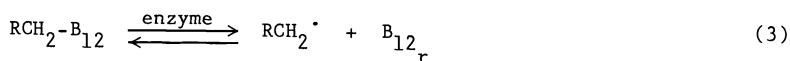
INTRODUCTION

Coenzyme B₁₂ (5'-deoxyadenosylcobalamin, abbreviated RCH₂-B₁₂), whose structure is depicted in Fig. 1, serves as a cofactor for a variety of enzymatic reactions, a common feature of which involves the 1,2-interchange of a hydrogen atom and another group (X = OH, NH₂, CH(NH₂)COOH, C(=CH₂)COOH, etc.) on adjacent carbon atoms according to eq. 1 (Ref. 1-3). A specific example of such a reaction is the deamination of ethanolamine catalyzed by ethanolamine ammonia lyase according to eq. 2 (Ref. 4).



Fig. 1. Coenzyme B₁₂

A widely accepted mechanism of such coenzyme B₁₂-dependent rearrangements, supported by a variety of studies on the enzymatic processes as well as on model systems, is depicted by eq. 3 and 4 (Ref. 1-3).



This mechanism encompasses the following sequence of steps: (i) enzyme-induced homolytic dissociation of the cobalt-carbon bond to generate cob(II)-alamin (i.e., vitamin B_{12r}) and a 5'-deoxyadenosyl radical (abbreviated CHR₂[·]), (ii) H-atom abstraction from the substrate to generate a substrate radical and 5'-deoxyadenosine (RCH₃), (iii) rearrangement of the resulting substrate radical (through a mechanism that is not fully understood and that probably differs from substrate to substrate) and (iv) abstraction of an H-atom from RCH₃ by the rearranged radical to complete the rearrangement reaction.

While the possible involvement of the cobalt complex in the substrate radical rearrangement step has been suggested, the evidence for this is inconclusive. At this stage it appears that the principal, if not only, role of the organometallic cofactor (i.e., of coenzyme B₁₂) in these reactions is to serve as a precursor for an organic free radical.

Model system studies have played, and promise to continue to play, several important roles in the study and understanding of such coenzyme B₁₂-dependent rearrangements, notably (Ref. 1):

1. In the development of methods of determining cobalt-alkyl bond dissociation energies, including that of coenzyme B₁₂, and in the elucidation of

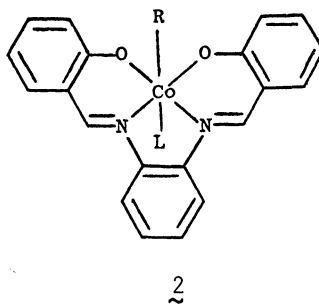
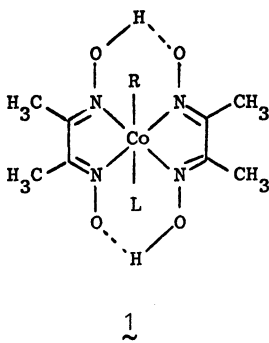
the factors that influence such bond dissociation energies and that may contribute to the weakening and dissociation of the cobalt-carbon bond under the conditions of the enzymatic reactions.

2. In achieving an understanding of the mechanisms of rearrangement of the substrate radical intermediates.

The present paper is concerned particularly with the first of the above themes.

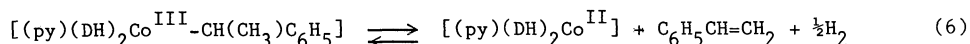
DETERMINATION OF COBALT-ALKYL BOND DISSOCIATION ENERGIES

The role of coenzyme B₁₂, encompassed by the mechanistic interpretation of eq. 3 and 4, implies a very weak cobalt-carbon bond. A troublesome feature of this interpretation has been the absence of direct supporting evidence that the cobalt-carbon bond in coenzyme B₁₂ (whose dissociation energy has not yet been determined) is sufficiently weak, that facile homolysis under the mild conditions of the enzymatic reactions is a plausible process. Indeed, hardly any cobalt-alkyl (or other transition metal-alkyl) bond dissociation energies were known reliably, nor were general methods for the determination of such bond dissociation energies (defined as the enthalpies of the process depicted by eq. 5) available, until quite recently (Ref. 5). Accordingly, as detailed below, we have undertaken the development of such methods and have successfully applied them to the determination of cobalt-alkyl bond dissociation energies of organocobalt compounds related to coenzyme B₁₂, notably alkyl-bis(dimethylglyoximate)cobalt(III) compounds (1, abbreviated [R-Co(DH)₂L], where DH₂ = dimethylglyoxime) and alkyl (N,N'-disalicylidene-o-phenylenediamine)cobalt(III) compounds (2, abbreviated [R-Co(Saloph)L] or Co-R). Both classes of compounds have been widely invoked in a variety of contexts as coenzyme B₁₂ analogues (Ref. 1).



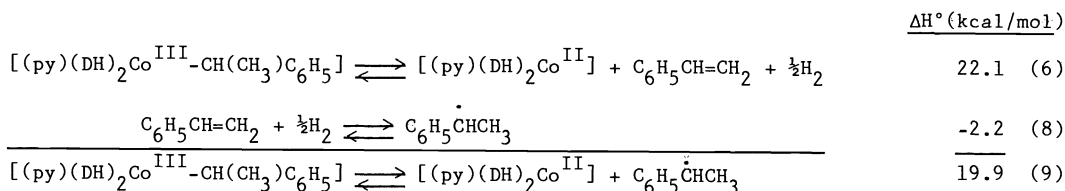
Equilibrium Method

We have found that for certain organocobalt compounds, exemplified by [(py)(DH)₂Co-CH(CH₃)C₆H₅] (py = pyridine) decomposition according to eq. 6 attains a measurable equilibrium under mild conditions (10°-40°C in solvents such as acetone or toluene) permitting the spectrophotometric determination of the equilibrium constant K₆ (1.3 x 10⁻⁵ M^{3/2} at 25°C) (Ref. 6). The temperature-dependence of K₆ yields the corresponding values of ΔH°₆ (22.1 kcal/mol) and ΔS°₆ (+52 cal/mol°K).



$$K_6 = \frac{[(\text{py})(\text{DH})_2\text{Co}^{\text{II}}][\text{C}_6\text{H}_5\text{CH}=\text{CH}_2][\text{H}_2]^{\frac{1}{2}}}{[(\text{py})(\text{DH})_2\text{Co}^{\text{III}}-\text{CH}(\text{CH}_3)\text{C}_6\text{H}_5]} \quad (7)$$

Using available data for the heats of formation of $\text{C}_6\text{H}_5\text{CH}=\text{CH}_2$ [$\Delta H_f^\circ(25^\circ\text{C}) = 35.2$ kcal/mol] (Ref. 7) and of the $\text{C}_6\text{H}_5\dot{\text{C}}\text{HCH}_3$ radical [$\Delta H_f^\circ(25^\circ\text{C}) = 33$ kcal/mol], (Ref. 8) the cobalt-carbon and dissociation energy of $[(\text{py})(\text{DH})_2\text{Co}-\text{CH}(\text{CH}_3)\text{C}_6\text{H}_5]$ (i.e., ΔH° for eq. 9) can be deduced to be 19.9 kcal/mol using the following thermochemical cycle:



It should be noted that this determination of the cobalt-carbon bond dissociation energy rests entirely upon thermodynamic considerations and is independent of the mechanism of reaction 6.

Values of the thermodynamic parameters and Co-C bond dissociation energies for various $[\text{L}(\text{DH})_2\text{Co}-\text{CH}(\text{CH}_3)\text{C}_6\text{H}_5]$ compounds, containing different axial ligands L, determined by this procedure, are listed in Table 1 (Ref. 9).

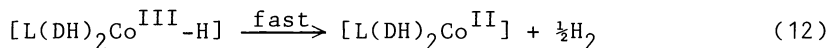
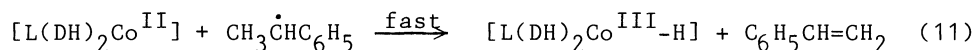
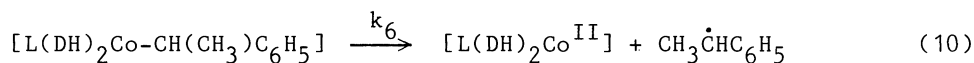
TABLE 1. Equilibrium and kinetic data for the decomposition of some $[\text{L}(\text{DH})_2\text{Co}-\text{CH}(\text{CH}_3)\text{C}_6\text{H}_5]$ compounds in acetone according to Eq. 6 (Ref. 9)

L	$10^6 K_6(25^\circ\text{C})$ $\text{M}^{3/2}$	ΔH_6° kcal/mol	ΔS_6° cal/mol $^\circ\text{K}$	$D_{\text{Co-R}}$ kcal/mol	$10^4 k_6(25^\circ\text{C})$ sec^{-1}	ΔH_6^\ddagger kcal/mol	ΔS_6^\ddagger cal/mol $^\circ\text{K}$
4-NH ₂ -Pyridine	5.5	23.4	54.3	21.2	4.0	23.1	3.8
4-CH ₃ -Pyridine	13.6	22.3	52.5	20.1	6.0	21.8	0.9
Pyridine	19.5	21.7	52.2	19.5	7.3	21.6	-0.2
4-CN-Pyridine	47.2	20.1	47.5	17.9	13.1	20.1	-3.9
Imidazole	4.0	23.0	52.5	20.8	1.7	23.0	1.9

Kinetic Method

The systems just described also provide a test of the validity of the kinetic approach to the determination of bond dissociation energies. Although the Co-C bond dissociation energies listed in Table 1 were deduced by a method that does not depend on the mechanism of reaction 6, a plausible mechanism encompasses the sequence of steps depicted by eq. 10-12, the rate-determining step being the homolysis of the Co-C bond. Kinetic measurements confirmed that these decomposition reactions obey first order kinetics, i.e., $-d[\text{L}(\text{DH})_2\text{Co}-\text{CH}(\text{CH}_3)\text{C}_6\text{H}_5]/dt = k_6[\text{L}(\text{DH})_2\text{Co}-\text{CH}(\text{CH}_3)\text{C}_6\text{H}_5]$, and yielded the values of ΔH_6^\ddagger and ΔS_6^\ddagger listed in Table 1. The finding that the values of ΔH_6^\ddagger are consistently about 2 kcal/mol higher than the corresponding Co-R bond dissociation energies lends strong support to this mechanistic interpretation

and implies that measurements of ΔH^\ddagger for reactions analogous to eq. 6 (or other reactions whose rates are determined by Co-C bond homolysis) can be used to deduce the corresponding Co-C bond dissociation energies in cases where the reactions do not achieve measurable equilibria.



Results of such kinetic measurements on other $[\text{L}(\text{DH})_2\text{Co}-\text{CH}(\text{CH}_3)\text{C}_6\text{H}_5]$ compounds, notably containing tertiary phosphine axial ligands, are listed in Table 2 (Ref. 11).

TABLE 2. Effect of axial ligands on the kinetics of decomposition of $[\text{L}(\text{DH})_2\text{Co}-\text{CH}(\text{CH}_3)\text{C}_6\text{H}_5]$ compounds in acetone according to eq. 6 (Ref. 10)

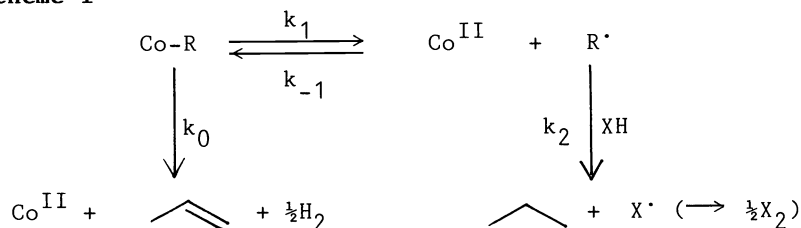
L	Cone Angle deg.	$10^3 k_6$ (25°C) sec ⁻¹	ΔH_6^\ddagger kcal/mol	ΔS_6^\ddagger cal/mol°K	$D_{\text{Co-R}}$ kcal/mol
$\text{P}(\text{CH}_3)_2\text{C}_6\text{H}_5$	122	0.10	25.9	10.5	24
$\text{P}(\text{CH}_2\text{CH}_2\text{CN})_3$	132	3.1	22.1	4.5	20
$\text{P}(\text{n-C}_4\text{H}_9)_3$	132	1.2	22.8	5.1	21
$\text{P}(\text{CH}_3)(\text{C}_6\text{H}_5)_2$	136	1.4	-	-	-
$\text{P}(\text{C}_2\text{H}_5)(\text{C}_6\text{H}_5)_2$	140	3.3	21.3	1.9	19
$\text{P}(\text{C}_6\text{H}_5)_3$	145	19.2	19.3	-1.4	17
$\text{P}(\text{cyclo-C}_6\text{H}_{11})_3$	170	480	-	-	-

The kinetic approach also has been applied to the determination of cobalt-alkyl bond energies in some organocobalt Schiff base compounds, $[\text{R}-\text{Co}(\text{Saloph})\text{L}]$ (2, abbreviated Co-R, L = pyridine) (Ref. 11). Such compounds were found to undergo thermal decomposition at conveniently measurable rates in pyridine solution at temperatures below 100°C in the presence of efficient radical traps such as the hydrogen donor, $\text{n-C}_8\text{H}_{17}\text{SH}$ (abbreviated XH). When R = n-propyl or isopropyl (i.e., an alkyl containing a β -hydrogen atom) the reaction yielded a mixture of propene and propane, exhibiting the kinetics (eq. 13) and product distribution (eq. 14) corresponding to Scheme I where $\text{CoR} = (\text{py})(\text{Saloph})\text{Co}-\text{C}_3\text{H}_7$ and $\text{Co}^{\text{II}} = (\text{py})(\text{Saloph})\text{Co}^{\text{II}}$ (Ref. 11).

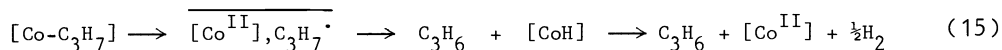
$$\frac{-d\ln[\text{CoR}]}{dt} = k_{\text{obsd}} = k_0 + \frac{k_1 k_2 [\text{XH}]}{k_{-1} [\text{Co}^{\text{II}}] + k_2 [\text{XH}]} \quad (13)$$

$$\frac{[\text{CH}_2=\text{CHCH}_3]}{([\text{CH}_3\text{CH}_2\text{CH}_3] + [\text{CH}_2=\text{CHCH}_3])} = k_0/k_{\text{obsd}} \quad (14)$$

Scheme I



The probable mechanism of the olefin-elimination step corresponding to k_0 is that depicted by eq. 15, i.e., β -hydrogen transfer between the $\text{C}_3\text{H}_7^\cdot$, Co^{II} geminate radical pair, followed by rapid decomposition of the resulting cobalt-hydride. Such β -hydrogen abstraction has been shown to be fast in closely related systems and, in one case, to occur within the cage lifetime of such a radical pair (Ref. 12). In any event, since k_0 is small compared with k_1 (in no case greater than ca 3%) its interpretation does not seriously affect the interpretation of the major k_1 -derived radical pathway. For R = neopentyl or benzyl (i.e., lacking a β -hydrogen atom) the olefin-producing path was absent. Accordingly, the organic products were exclusively neopentane and toluene, and the kinetics conformed to eq. 13 with $k_0 = 0$.



The results of these kinetic measurements are summarized in Table 3 (Ref. 11) together with the Co-R bond dissociation energies ($D_{\text{Co-R}}$), deduced from ΔH_1^\ddagger on the assumption that recombination of Co^{II} and R^\cdot is diffusion controlled, i.e., that $\Delta H_{-1}^\ddagger \sim 2$ kcal/mol, hence $D_{\text{Co-R}} \sim \Delta H_1^\ddagger - 2$ kcal/mol.

All the Co-alkyl bond dissociation energies determined in these studies, both for $[\text{R-Co}(\text{DH})_2\text{L}]$ compounds (Tables 1 and 2) and $[\text{R-Co}(\text{Saloph})\text{L}]$ compounds (Table 3), lie in the range 17-25 kcal/mol. Comparable values have recently been estimated for some alkylcobalamins by a similar kinetic procedure (Ref. 13). These values lie in an appropriately low range to be consistent with, and supportive of, the proposed role of Co-C bond homolysis in coenzyme B_{12} -promoted reactions according to eq. 3 and 4. Co-C bond homolysis probably also is involved in other reactions of organocobalt compounds, for example the photochemical or thermal insertion of O_2 according to eq. 16 (Refs. 14 and 15).

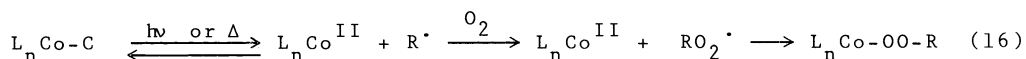


TABLE 3. Kinetic data for the decomposition of some $[\text{R-Co}(\text{Saloph})\text{L}]$ compounds according to Scheme I (Ref. 11)

R	$k_0(70^\circ\text{C})$ sec ⁻¹	ΔH_0^\ddagger kcal/mol	ΔS_0^\ddagger cal/mol ^o K	$k_1(70^\circ\text{C})$ sec ⁻¹	ΔH_1^\ddagger kcal/mol	ΔS_1^\ddagger cal/mol ^o K	$(k_{-1}/k_2)_{70^\circ\text{C}}$	$D_{\text{Co-R}}$ kcal/mol
$-\text{CH}_2\text{CH}_2\text{CH}_3$	1.0×10^{-5}	23.4	-15.1	4.7×10^{-4}	27.1	2.6	10	25
$-\text{CH}_2(\text{CH}_3)_2$	1.9×10^{-3}	19.8	-15.5	5.7×10^{-2}	21.8	-2.9	93	20
$-\text{CH}_2\text{C}(\text{CH}_3)_3$	-	-	-	3.4×10^{-2}	20.3	-6.2	8	18
$-\text{CH}_2\text{C}_6\text{H}_5$	-	-	-	1.2×10^{-2}	23.6	1.3	70	22

EFFECTS OF ELECTRONIC AND STERIC PARAMETERS ON COBALT-ALKYL BOND DISSOCIATION ENERGIES

The Co-C bond dissociation energies listed in Table 1 span the range 17.9 to 21.2 kcal/mol. For the series of complexes containing *p*-substituted pyridines as the axial ligands, for which the steric influences presumably are constant, the Co-C bond dissociation energy increases with the basicity of the axial ligand L according to the trend depicted in Figure 2. This is not unexpected since dissociation of the Co-C bond according to eq. 9 involves a decrease in the formal oxidation state of cobalt from +3 to +2. By favoring the higher oxidation state more basic ligands should, accordingly, stabilize the organocobalt compound and reduce the driving force for Co-C bond homolysis. Analogous reasoning has been invoked to explain the increase in reactivity of Co^{III}(DH)₂L complexes toward organic halides with increasing basicity of L (Ref. 16).

The influence of steric factors on Co-C bond dissociation energies is illustrated in Table 2 and Fig. 3. When L is a tertiary phosphine ligand the value of k_6 was found to increase markedly (and the value of ΔH_6^\ddagger , and hence of $D_{\text{Co-R}}$, to decrease correspondingly) with the cone angle (Ref. 17) of L, e.g., from $1.0 \times 10^{-4} \text{ sec}^{-1}$ at 25°C ($\Delta H_6^\ddagger = 26 \text{ kcal/mol}$, $D_{\text{Co-R}} \sim 24 \text{ kcal/mol}$) for L = dimethylphenylphosphine (cone angle 122°), through $1.9 \times 10^{-2} \text{ sec}^{-1}$ ($\Delta H_6^\ddagger = 19 \text{ kcal/mol}$, $D_{\text{Co-R}} \sim 17 \text{ kcal/mol}$) for L = triphenylphosphine (cone angle 145°) to ca 1.5 sec^{-1} for L = tricyclohexylphosphine (cone angle 170°).

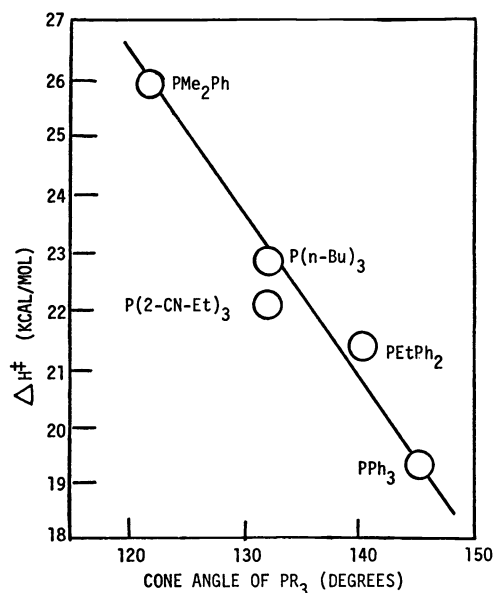
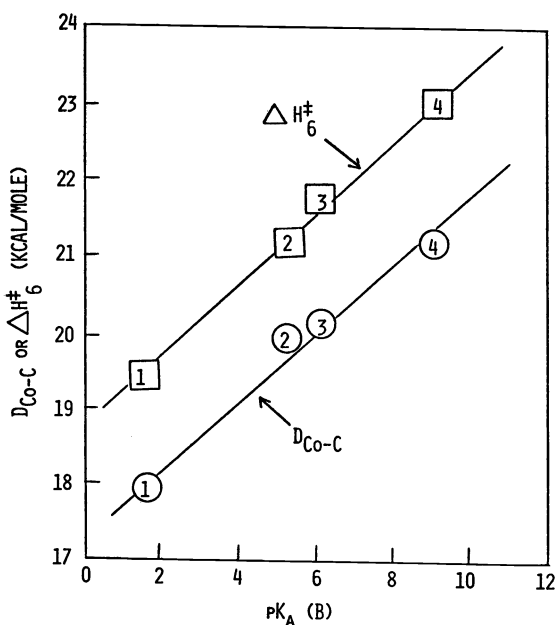


Fig. 2. Dependence of $D_{\text{Co-C}}$ and ΔH_6^\ddagger on pK_a of L. Fig. 3. Effect of the cone angle of the axial phosphine ligand (L) on ΔH_6^\ddagger for the decomposition of $[\text{L}(\text{DH})_2\text{Co}-\text{CH}(\text{CH}_3)\text{C}_6\text{H}_5]$ (data from Table 2).

Marked steric influences also are apparent in the trend of $D_{\text{Co-R}}$ bond dissociation energies of (py)(Saloph)Co-R compounds listed in Table 3. These $D_{\text{Co-R}}$ values span the range 18-25 kcal/mol. While the trend, in part (e.g., *n*-propyl > benzyl), parallels that for other alkyl bond dissociation energies (e.g., R-H), the low values of $D_{\text{Co-R}}$ for isopropyl and neopentyl, compared with benzyl, attest to the importance of steric effects in such cobalt complexes. In this context our results suggest that neopentyl is at least as sterically demanding as isopropyl.

The influence of steric factors also is reflected in the results of structural studies on $[\text{L}(\text{DH})_2\text{Co-R}]$ compounds. Such results, summarized in Table 4, reveal significant lengthening of the Co-C bond (as well as other distortions reflecting steric hindrance) with increasing steric bulk of either the R or L group (Ref. 18).

TABLE 4. Selected Co-C and Co-L bond lengths in $[L(DH)_2Co-R]$ compounds (Ref. 18)

R	L	Co-C (Å)	Co-L (Å)
CH ₃	H ₂ O	1.990	2.058
"	Pyridine	1.998	2.068
"	P(CH ₃) ₃	2.015	2.294
"	P(C ₆ H ₅) ₃	2.026	2.418
CH ₂ C(CH ₃) ₃	H ₂ O	2.044	2.056
"	Pyridine	2.060	2.081
"	P(CH ₃) ₃	2.084	2.316
"	P(C ₆ H ₅) ₃	2.118	2.460

COMPARISON OF DIFFERENT ORGANOCOBALT COMPOUNDS AND EXTENSION TO ORGANOCOBALAMINS

The approaches described above have not been applied successfully to the determination of cobalt-carbon bond dissociation energies in cobalamins, although the prospects of accomplishing this would appear to be promising. Nevertheless, some inferences about cobalt-carbon bond dissociation energies in cobalamins can be drawn from the results obtained on other organocobalt compounds. Comparisons of the redox properties of various cobalt complexes, and of the reactivities of various cobalt(II) complexes (including vitamin B₁₂^r) toward organic halides suggests that cobalamins fall somewhere between the dimethylglyoxime and Schiff base cobalt complexes discussed in this paper (Ref. 1). Comparison of the corresponding Co-H bond dissociation energies suggests a similar trend, as does comparison of the stabilities of the corresponding cobalt-benzyl compounds (benzyl cobalamin being significantly less stable than the Co(DH)₂-benzyl analogues) (Ref. 1). On the basis of such comparisons it seems likely that the 5'-deoxyadenosyl-cobalt bond dissociation energy in coenzyme B₁₂ is in the range of cobalt-alkyl bond dissociation energies that we have determined for model compounds, notably of the Co(Saloph)-R compounds, i.e., 18-25 kcal/mol.

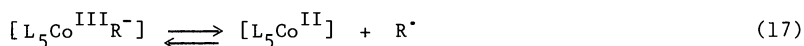
FACTORS CONTRIBUTING TO COBALT-CARBON BOND WEAKENING AND DISSOCIATION

Possible causes of enzyme-induced cobalt-carbon bond weakening and bond dissociation include: (1) axial ligand substitution, i.e., displacement of the 5,6-dimethylbenzimidazole ligand of coenzyme B₁₂ by another ligand (e.g., a sulfur-bonded cysteine residue) which weakens the cobalt-carbon bond through electronic or steric influences, (2) oxidation or reduction of the coenzyme which has been shown to induce cobalt-carbon bond dissociation in model organocobalt compounds (Ref. 1) and (3) conformational distortion of the corrin ring resulting in sterically unfavorable bending of the latter upward toward the 5'-deoxyadenosyl group. The influences of steric factors on cobalt-carbon bond dissociation energies identified in the studies described in this paper, as well as other lines of evidence, favor the last of the above interpretations (Ref. 1).

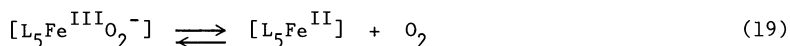
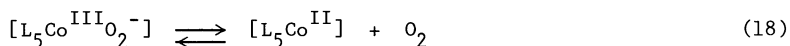
COMPARISONS WITH THE REVERSIBLE BINDING OF DIOXYGEN

The weakness of the cobalt-carbon bonds in coenzyme B₁₂ and related organocobalt compounds (collectively denoted as $[L_2Co^{III}R]$) permits homolytic dissociation of such bonds to occur under mild conditions (below 100°C).

Because recombination of the resulting cobalt(II) complex and free radical generally is rapid such dissociation processes, depicted by eq. 17, are readily reversible. Such processes also can be described as (inner sphere) redox changes and, as already noted, the influence of electronic factors on cobalt-carbon bond dissociation energies (Table 1 and Fig. 2) can be interpreted in terms of the accompanying changes in the oxidation state of the cobalt (eq. 17).



There is at least a formal parallel between the process depicted by eq. 17 and the reversible binding of dioxygen by cobalt or iron complexes (e.g., myoglobin) in accord with eq. 18 and 19.



This parallel is quite far-reaching and is reflected in trends in the dependence of Co-O₂ (and presumably Fe-O₂) bond dissociation free energies and enthalpies on electronic factors that parallel those of Co-R bond dissociation trends (Table 5 and Fig. 4) (Refs. 19 and 20). Indeed, typical Co-O₂ bond dissociation energies in such reversible oxygen carriers lie in the same range (10-20 kcal/mol) as the values of the Co-R bond dissociation energies that we have determined.

In the light of these considerations it is not unreasonable to describe the role of coenzyme B₁₂ in biological systems as that of a "reversible free radical carrier" analogous to the role of myoglobin or hemoglobin as a "reversible oxygen carrier." Thus, coenzyme B₁₂ fulfills its biochemical role by serving as a "free radical reservoir," from which (5'-deoxyadenosyl) free-radicals are reversibly released under mild conditions, just as oxy-hemoglobin serves as a reservoir for the storage and reversible release of dioxygen. Significant questions, in this context, relate to the alternative choices of cobalt and iron, as well as of the corrin and porphyrin ligand systems, for these two parallel functions.

TABLE 5. Effect of axial ligands (L) on the reversible binding of O₂ to protoporphyrin IX dimethyl ester cobalt(II) [LCo^{II}(P)] and N,N'-ethylenbis(benzoylacetiminato)cobalt(II) [LCo^{II}(benacen)] in toluene

Co Complex	L	pK _a of L	Log K _{O₂} mm ⁻¹	ΔH° kcal/mol	ΔS° cal/mol°K	Ref
[LCo ^{II} (P)]	4-CN-Py	1.8	-3.8 (-45°C)	-	-	19
"	Py	5.2	-2.84 "	-9.2	-53	19
"	4-t-Bu-Py	6.0	-2.77 "	-9.8	-56	19
"	4-NH ₂ -Py	9.1	-2.05 "	-9.9	-53	19
[LCo ^{II} (Benacen)]	4-CN-Py	1.8	-2.68 (-21°C)	-16.9	-77	20
"	Py	5.2	-2.03 "	-16.6	-75	20
"	3,4-(CH ₃) ₂ Py	6.4	-1.66 "	-16.8	-75	20

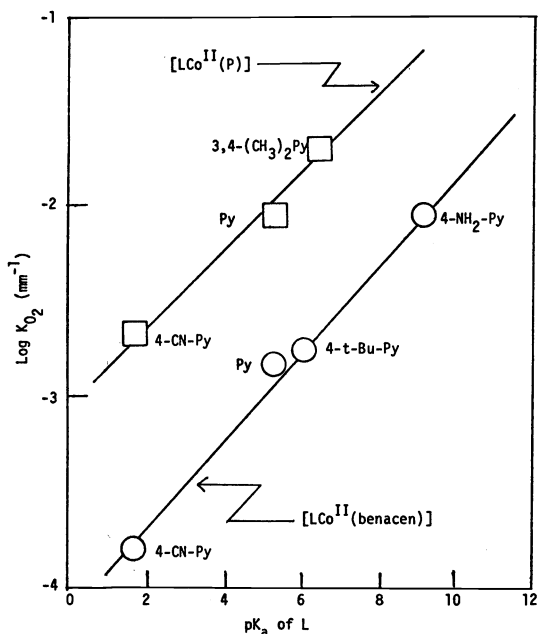


Fig. 4. Effect of axial ligands (L) on the reversible binding of O₂ to protoporphyrin IX ester cobalt (II) [LCo^{II}(P)] and N,N'-ethylenebis(benzoylacetonato)-cobalt (II) [LCo^{II}(benacen)] in toluene.

Acknowledgment - This research was supported by grants from the National Science Foundation and the National Institutes of Health.

REFERENCES

1. J. Halpern, in *B₁₂*, Vol. I, D. Dolphin, Ed., pp. 501-541, Wiley, New York (1982), and references therein.
2. B. M. Babior, *Acc. Chem. Res.*, **8**, 376-383 (1975).
3. R. H. Abeles and D. Dolphin, *Acc. Chem. Res.*, **9**, 114-120 (1976).
4. B. M. Babior, in *B₁₂*, Vol. II, D. Dolphin, Ed., Wiley, New York (1982) pp. 263-287.
5. J. Halpern, *Acc. Chem. Res.*, **15**, 238-244 (1982).
6. J. Halpern, F. T. T. Ng and G. L. Rempel, *J. Am. Chem. Soc.*, **101**, 7124-7126 (1979).
7. D. S. Stull, E. F. Westrum and G. C. Sinke, *The Chemical Thermodynamics of Organic Compounds*, Wiley, New York (1969).
8. J. A. Kerr, *Chem. Rev.*, **66**, 465-500 (1965).
9. F. T. T. Ng, G. L. Rempel and J. Halpern, *J. Am. Chem. Soc.*, **101**, 621-623 (1982).
10. F. T. T. Ng, G. L. Rempel and J. Halpern, unpublished results.
11. T. T. Tsou, M. Loots and J. Halpern, *J. Am. Chem. Soc.*, **104**, 623-624 (1982).
12. R. L. Sweany and J. Halpern, *J. Am. Chem. Soc.*, **99**, 8335-8337 (1977).
13. G. N. Schrauzer and J. Grate, *J. Am. Chem. Soc.*, **103**, 541-546 (1981).
14. (a) C. Giannotti, A. Gaudemer and C. Fontaine, *Tetrahedron Lett.*, 3209-3212 (1970); (b) K. N. V. Duong, C. Fontaine, M. C. Giannotti and A. Gaudemer, *Tetrahedron Lett.*, 1187-1189 (1971); (c) C. Fontaine, K. N. V. Duong, C. Merienne, A. Gaudemer and C. Giannotti, *J. Organomet. Chem.*, **38**, 167-178 (1972); (d) C. Giannotti, C. Fontaine and A. Gaudemer, *J. Organomet. Chem.*, **39**, 381-387 (1972).
15. F. R. Jensen and R. C. Kiskis, *J. Am. Chem. Soc.*, **97**, 5825-5831 (1975).
16. J. Halpern and P. Phelan, *J. Am. Chem. Soc.*, **94**, 1881-1886 (1972).
17. C. A. Tolman, *Chem. Rev.*, **77**, 313-348 (1977).
18. N. Bresciani-Pahor, M. Calligaris, G. Nardin and L. Randaccio, *J. Chem. Soc., Dalton Trans.*, 2549-2551 (1982), and references therein.
19. D. V. Stynes, H. C. Stynes, B. R. James and J. A. Ibers, *J. Am. Chem. Soc.*, **95**, 1796-1801 (1973).
20. M. J. Carter, D. P. Rillema and F. Basolo, *J. Amer. Chem. Soc.*, **96**, 392-400 (1974).